



# Phosphoproteomics of cardiac Nav1.5 channels



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# Phosphoproteomics of cardiac Nav1.5 channels

## Research experiences: Céline Marionneau

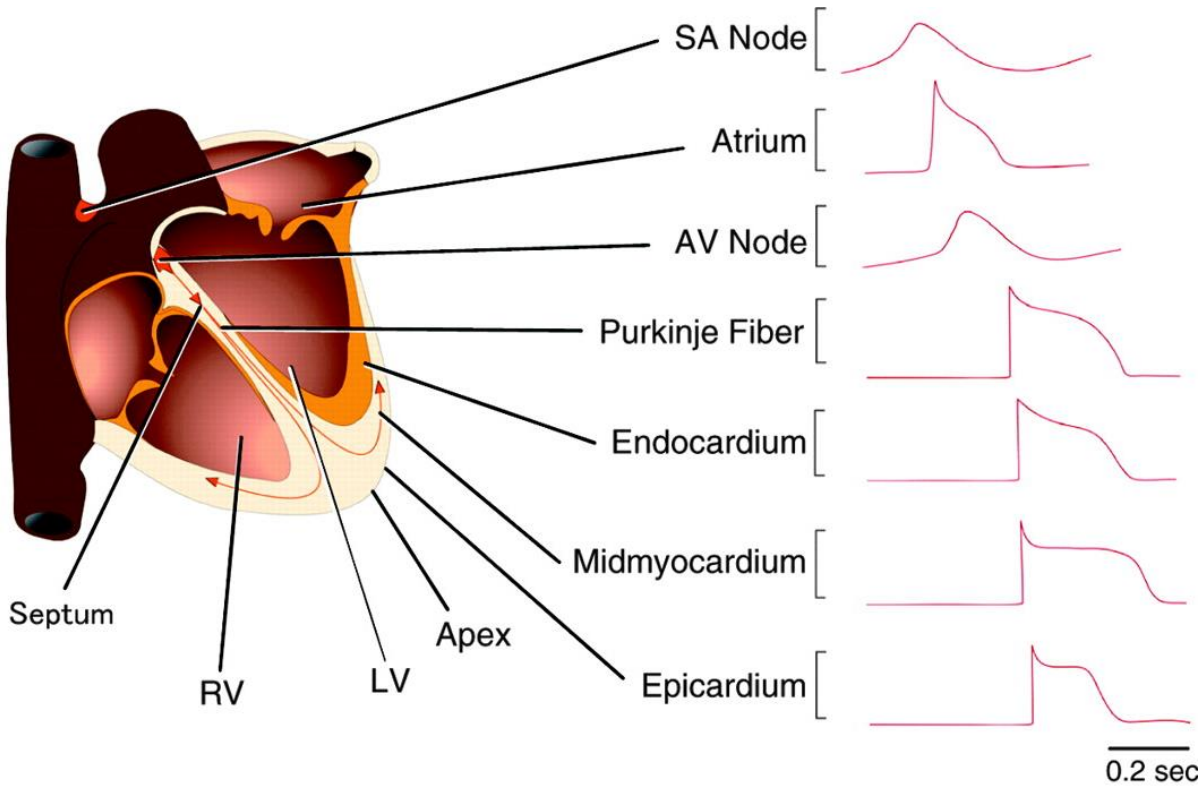
- 2001-2005: PhD (University of Nantes, *l'institut du thorax, Nantes*)  
Functional genomics of cardiac ion channels
- 2005-2009 : Postdoctoral fellow (Washington University, Saint Louis, MO, USA)  
Post-translational regulation of neuronal Kv4.2 channels
- 2009-2010 : Postdoctoral fellow (*l'institut du thorax, Nantes*)
- Since Nov 2010 : Chargée de recherche au CNRS (*Equipe II, l'institut du thorax, Nantes*)  
Post-translational regulation of cardiac Nav1.5 channels

# Phosphoproteomics of cardiac Nav1.5 channels

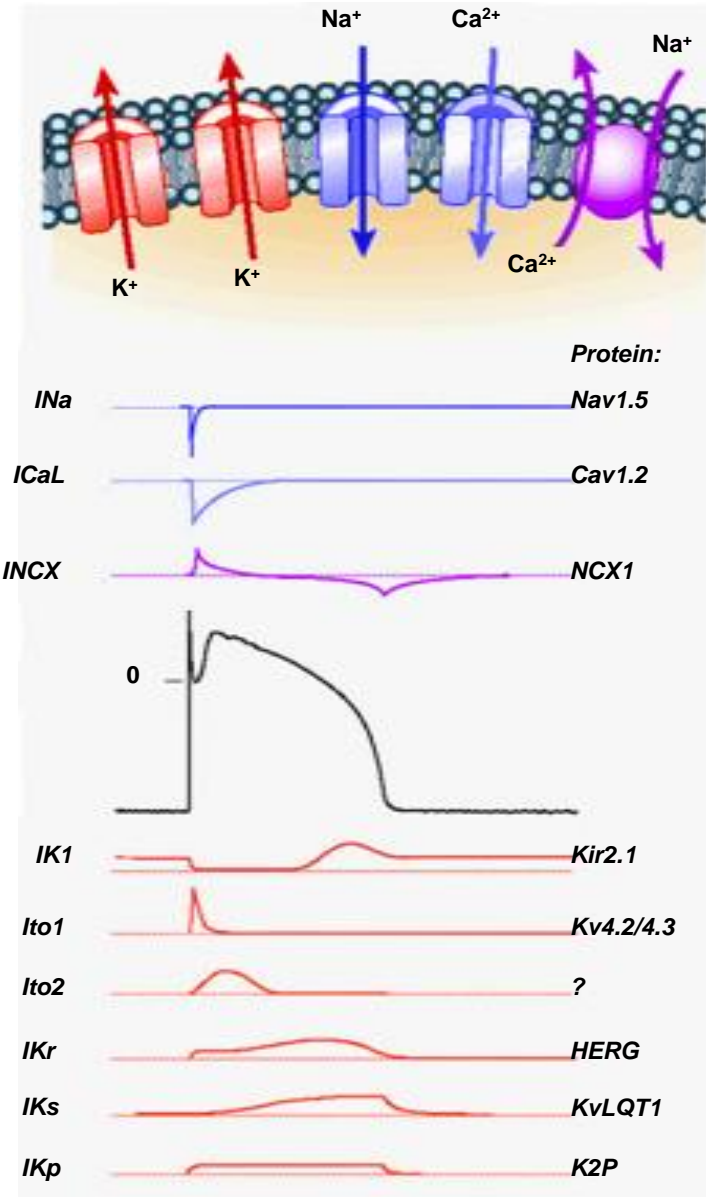
From this conference, you should know:

- Context : Cardiac electrophysiology, Nav1.5 channels
- Proteomics : goals, methods
- Apprehend the research developed

# Cardiac excitability

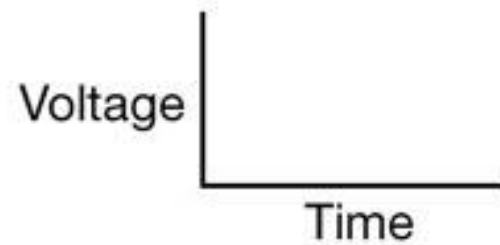
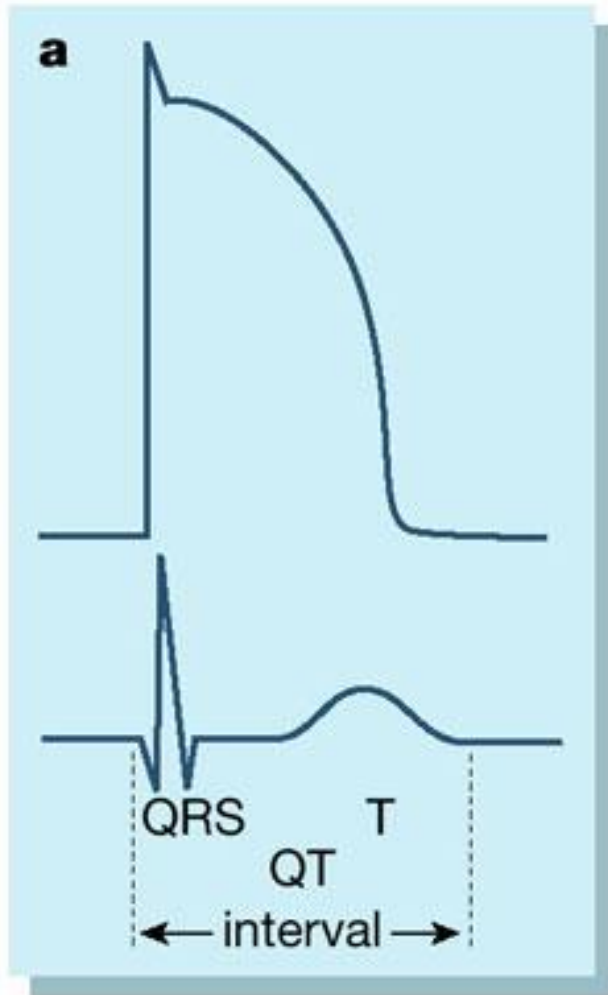


Nerbonne and Kass, *Physiol Rev* 2005

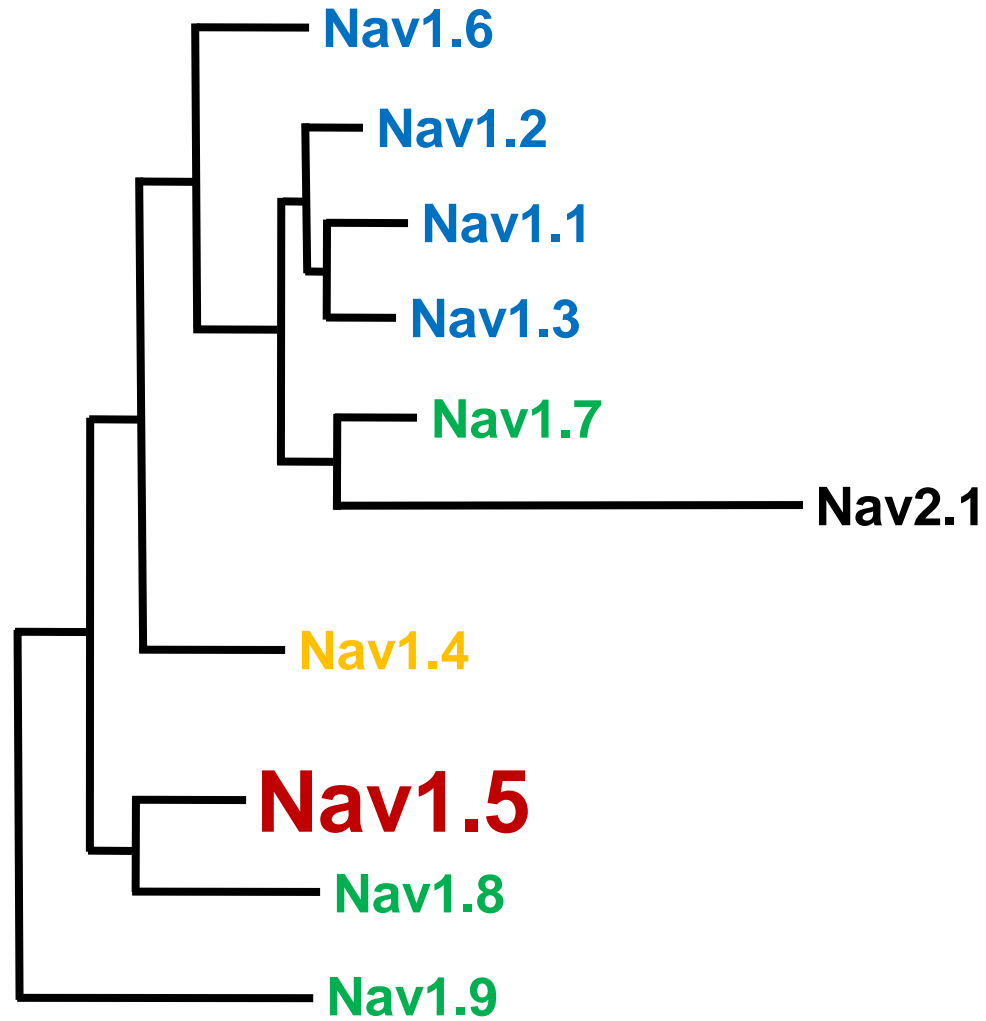
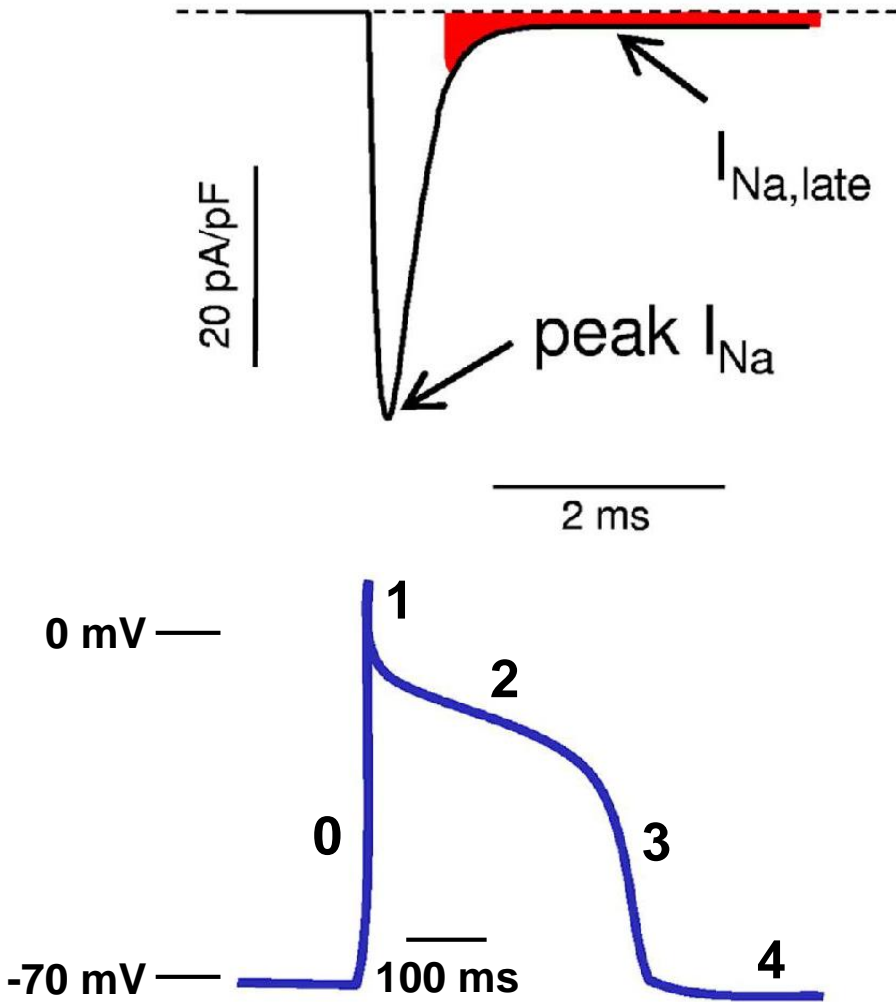


Marban, *Nature* 2002

# Electrocardiograms : ECG



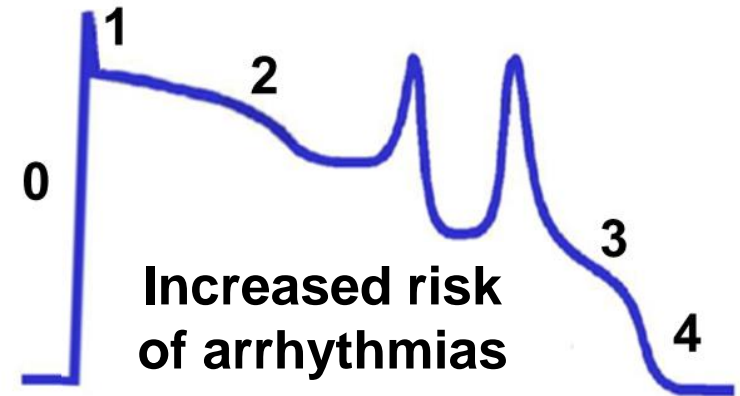
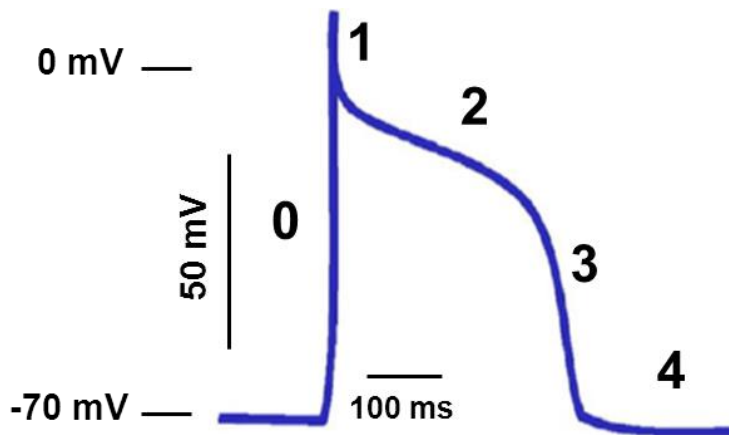
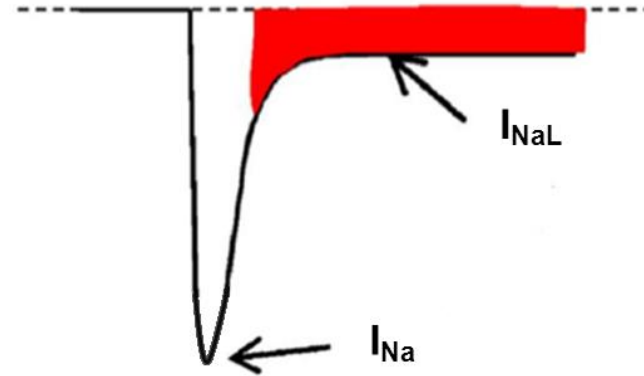
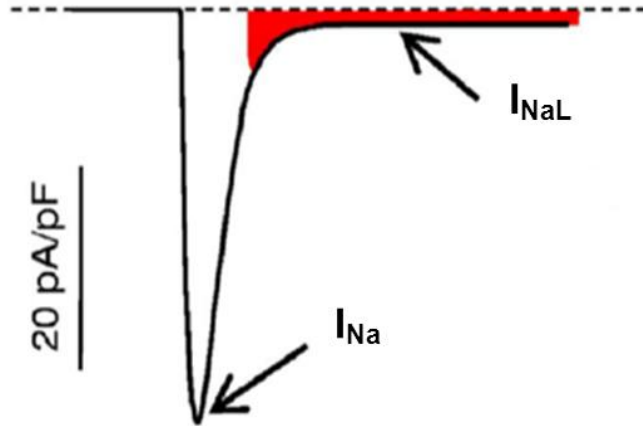
# Cardiac voltage-gated Na<sup>+</sup> channels



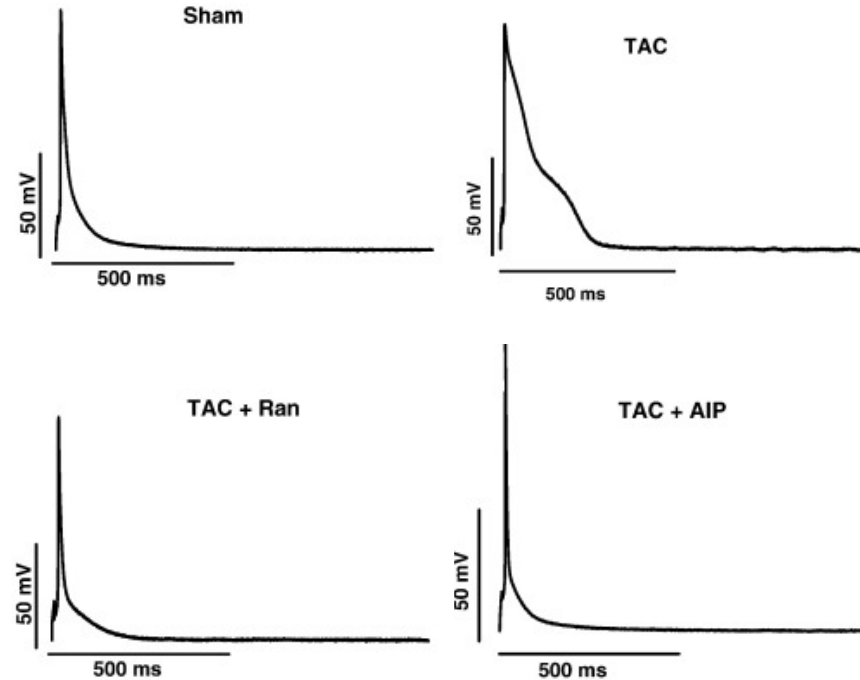
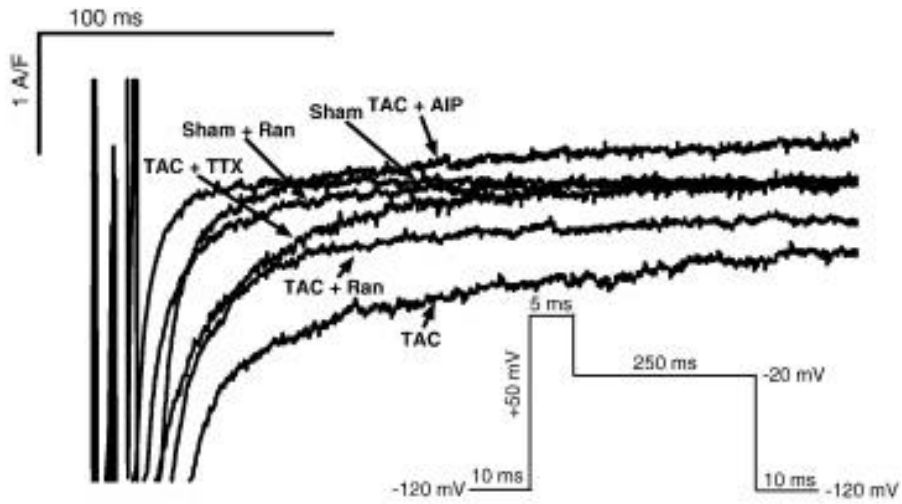
# Nav1.5 channels in heart failure

Normal

Disease

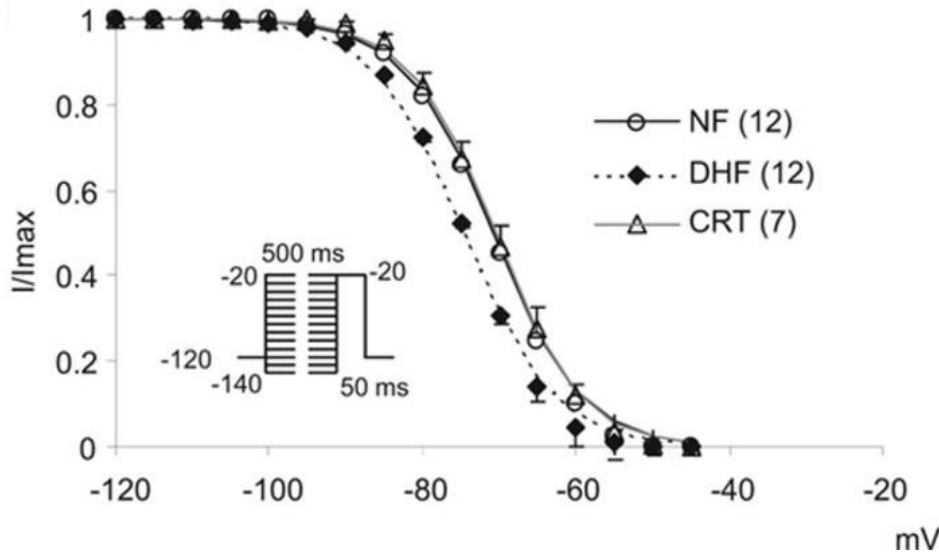


# CaMKII increases $I_{NaL}$ in heart failure

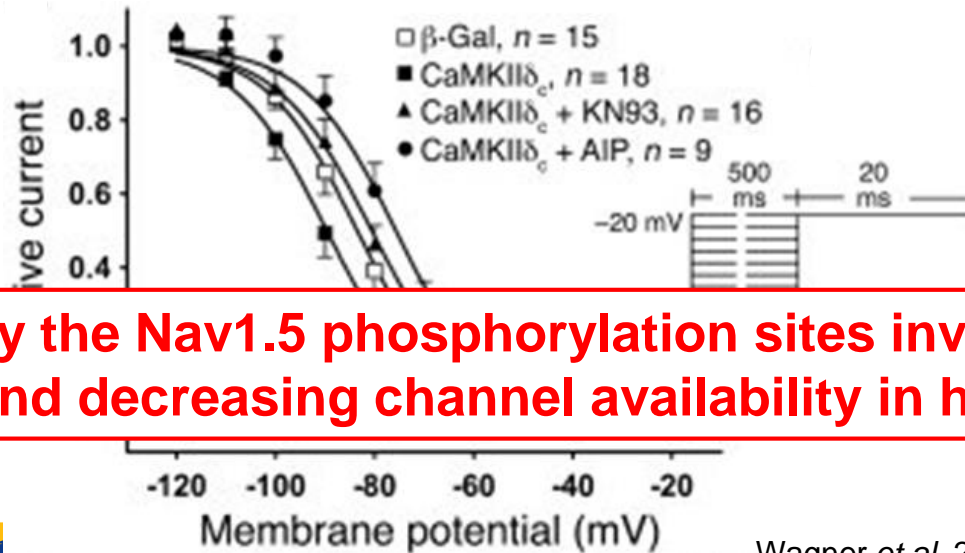
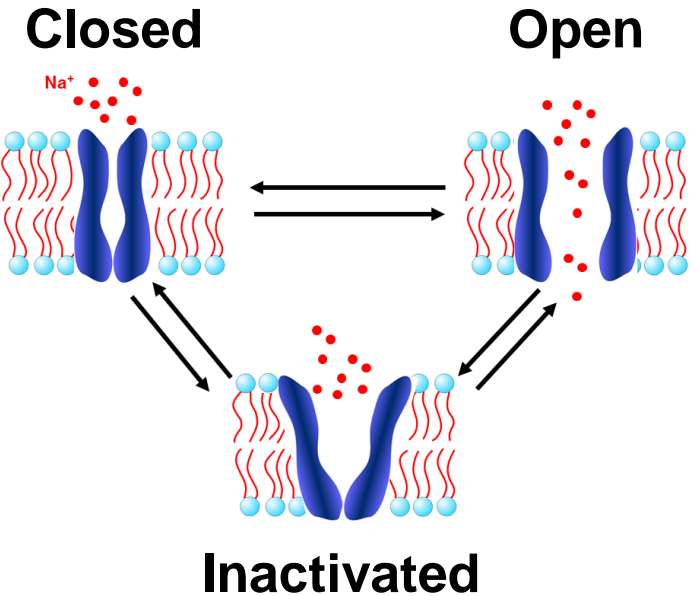




# CaMKII decreases Nav1.5 channel availability



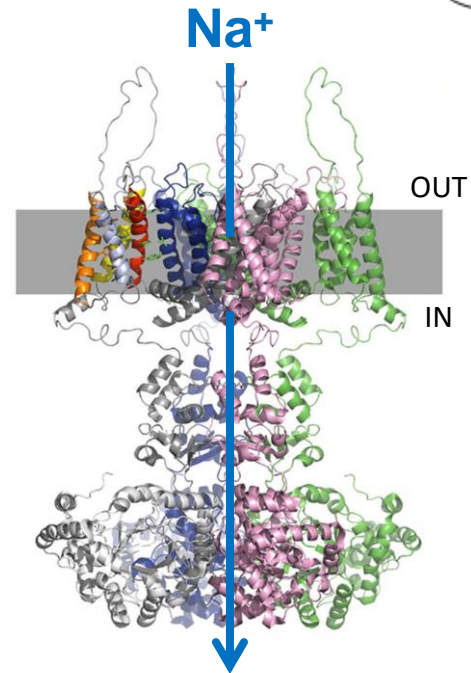
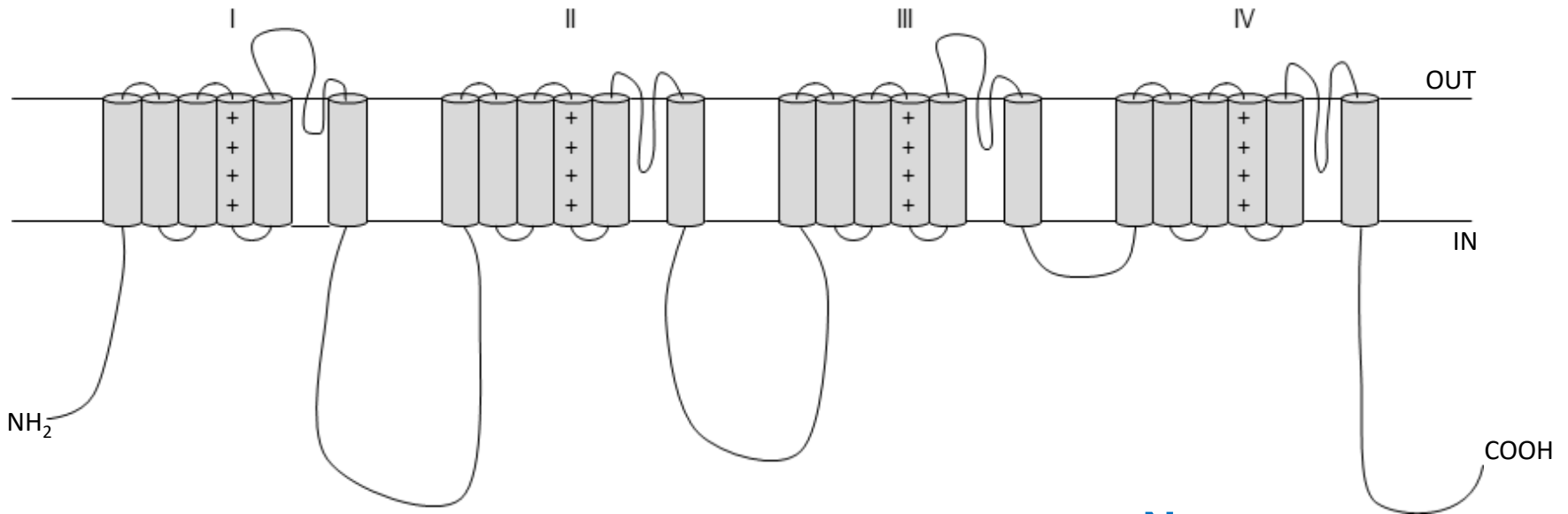
Aiba *et al*, 2013



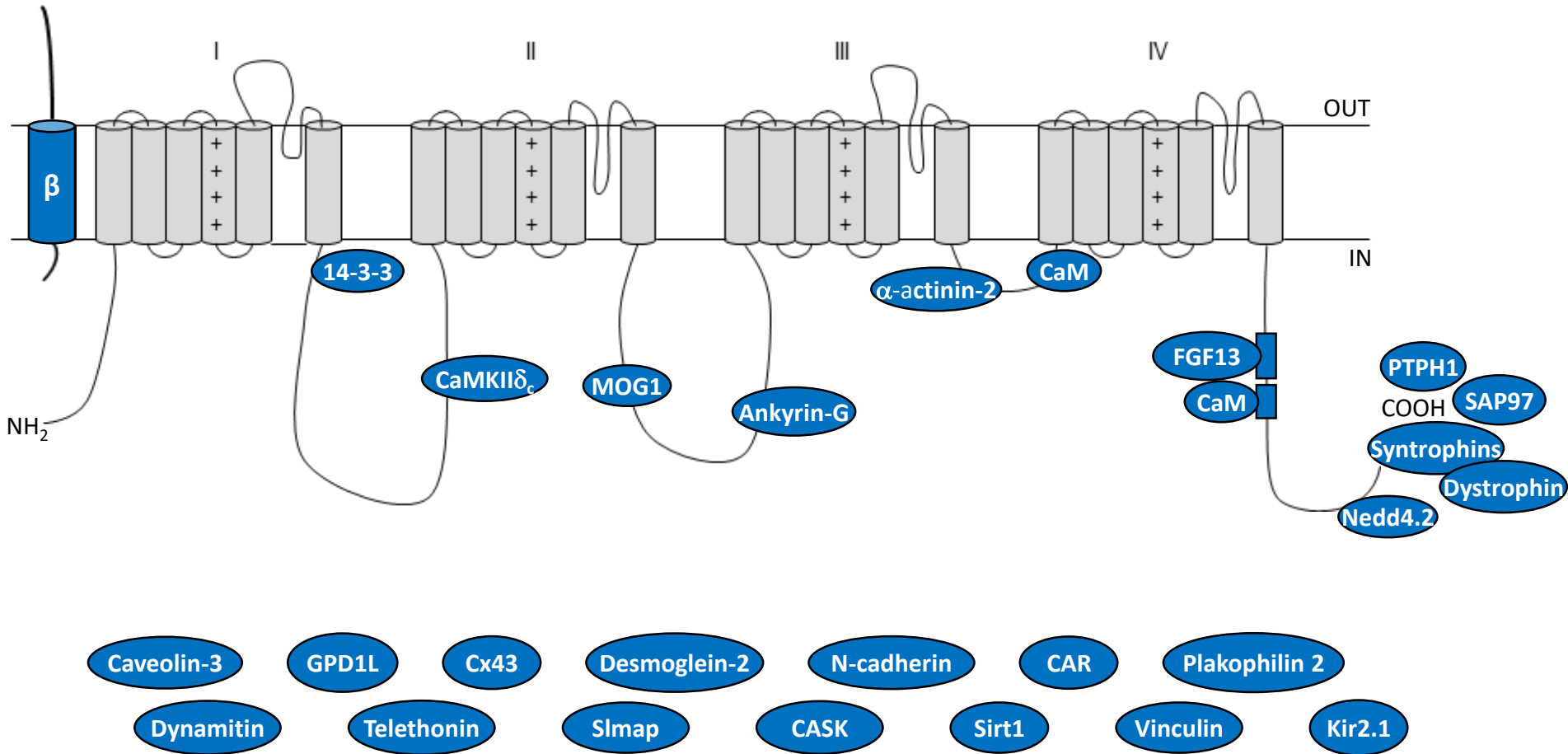
Wagner *et al*, 2006

**Goals: to identify the Nav1.5 phosphorylation sites involved in increasing  $I_{NaL}$  and decreasing channel availability in heart failure**

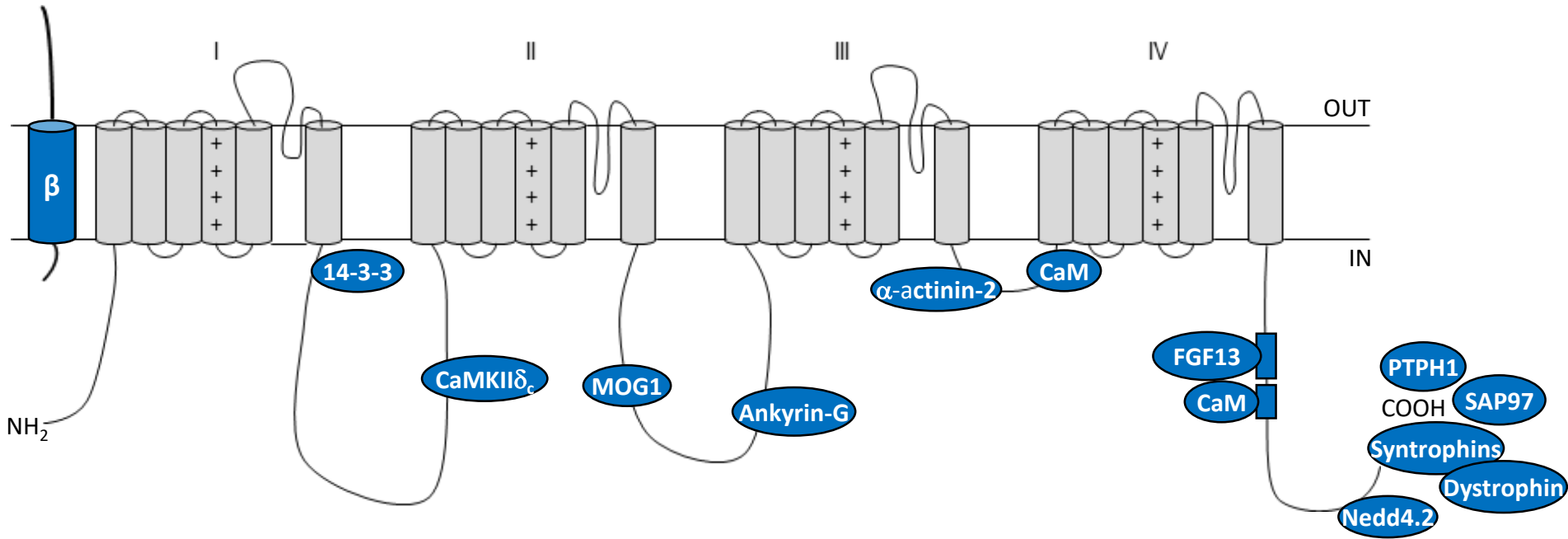
# Cardiac Nav1.5 channel complexes



# Cardiac Nav1.5 channel complexes



# Cardiac Nav1.5 channel complexes



O-GlcNAcylation

Glycosylation

Phosphorylation

Methylation

Acetylation

Oxidation

Nitrosylation

Lipoxidation

Ubiquitylation

Palmitoylation

SUMOylation

Caveolin-3

GPD1L

Cx43

Desmoglein-2

N-cadherin

CAR

Plakophilin 2

Dynamitin

Telethonin

Slmap

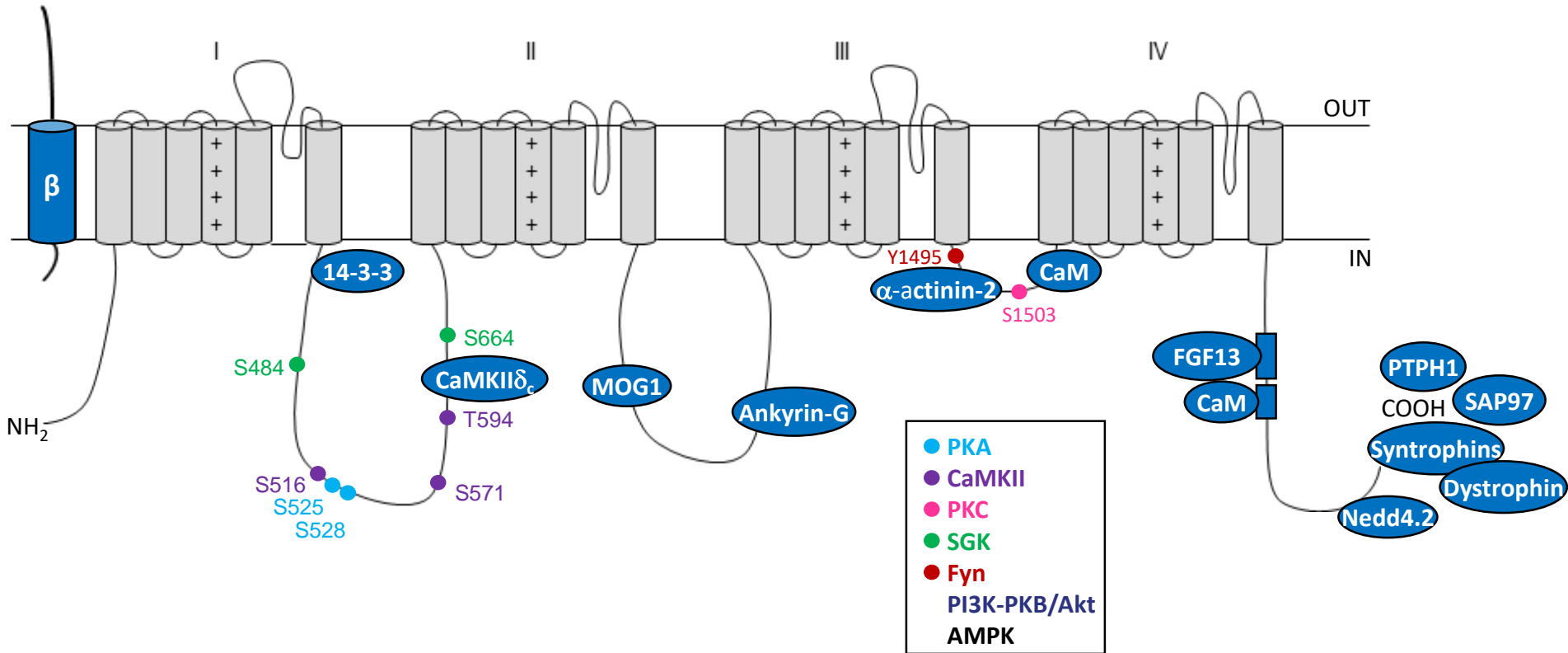
CASK

Sirt1

Vinculin

Kir2.1

# Before 2012 : 9 phosphorylation sites



O-GlcNAcylation

Glycosylation

Phosphorylation

Methylation

Acetylation

Oxidation

Nitrosylation

Lipoxidation

Ubiquitylation

Palmitoylation

SUMOylation

Caveolin-3

GPD1L

Cx43

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N-cadherin

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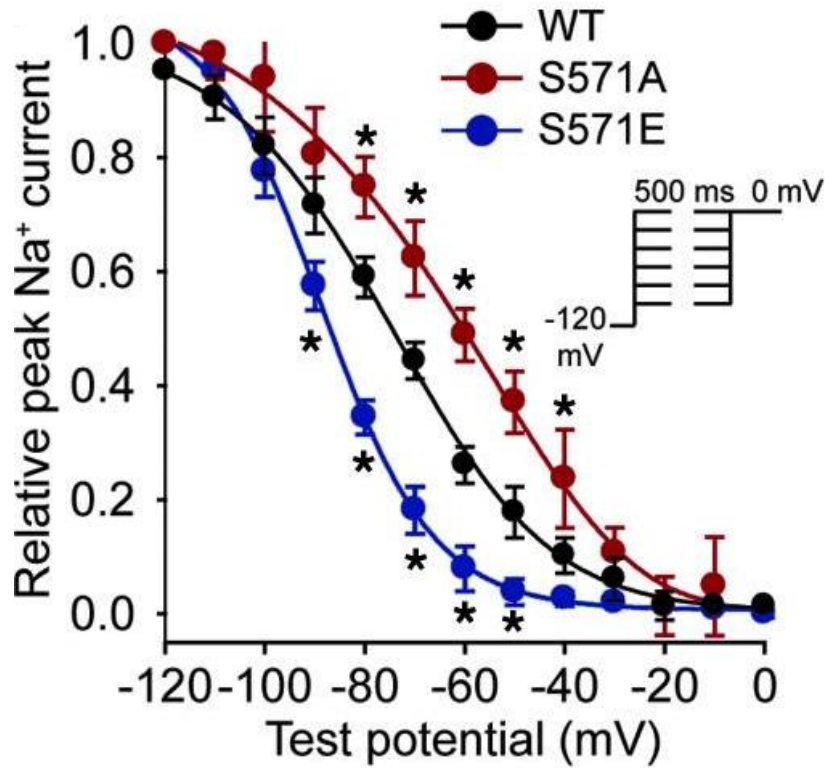


# Methods to identify phosphorylation sites

- *In silico analyses*
    - phosphorylation consensus sequence (CaMKII: RXX[S/T])
  - *In vitro analyses*
    - kinase assay + mass spectrometry
  - *In situ analyses*
    - purification of proteins + mass spectrometry
- ➔ Functional analyses :
- directed mutagenesis (Ser/Thr > Glu/Asp ; Ser/Thr > Ala)

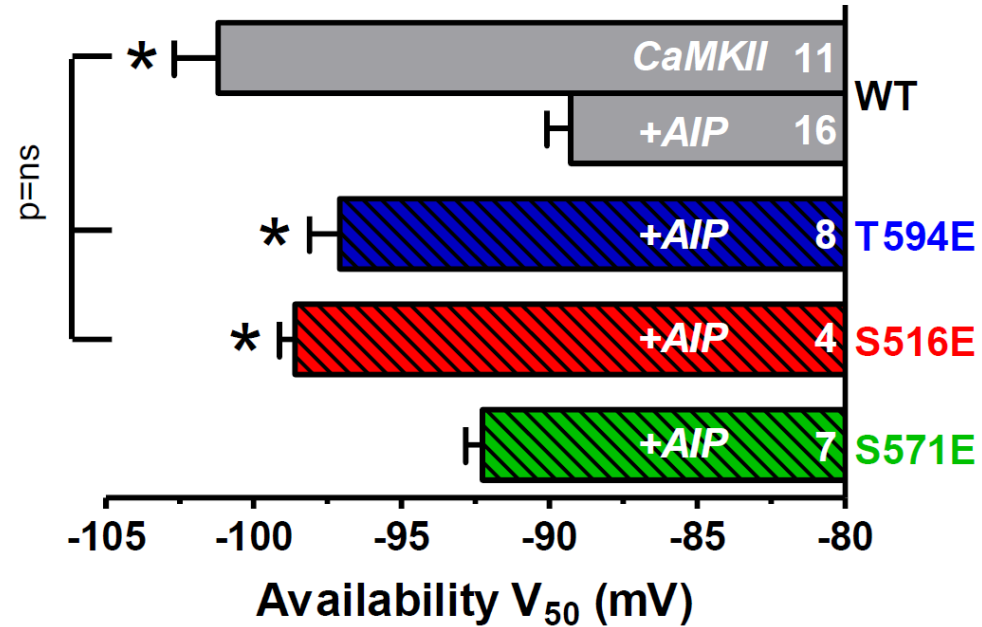
# CaMKII phosphorylation sites

*In silico*



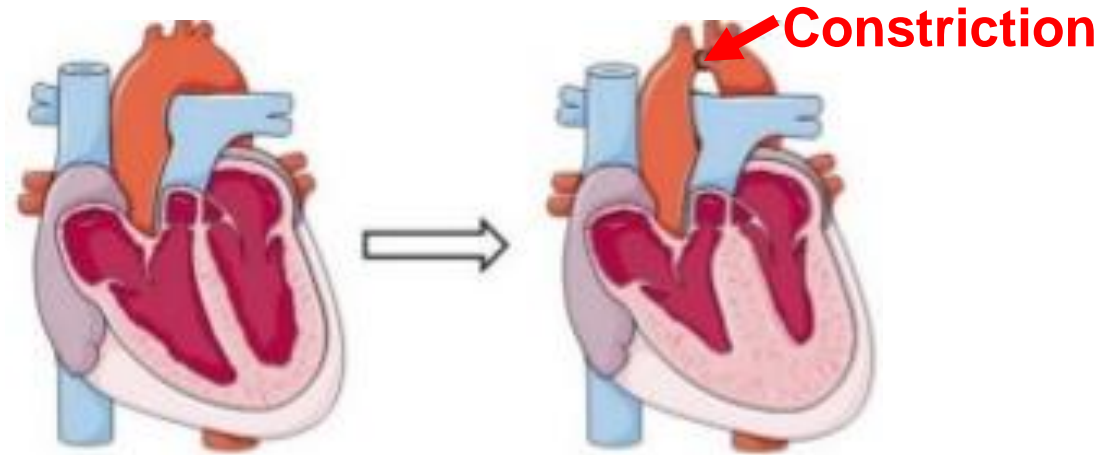
Hund *et al*, 2010

*In vitro*



Ashpole *et al*, 2012

# Phosphoproteomics of cardiac Nav1.5 channels

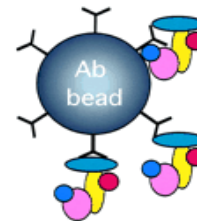
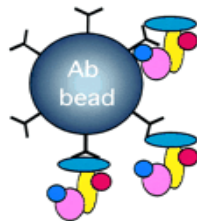


**Sham  
Ventricles**

**TAC  
Ventricles**

**m $\alpha$ NavPAN-IPs**

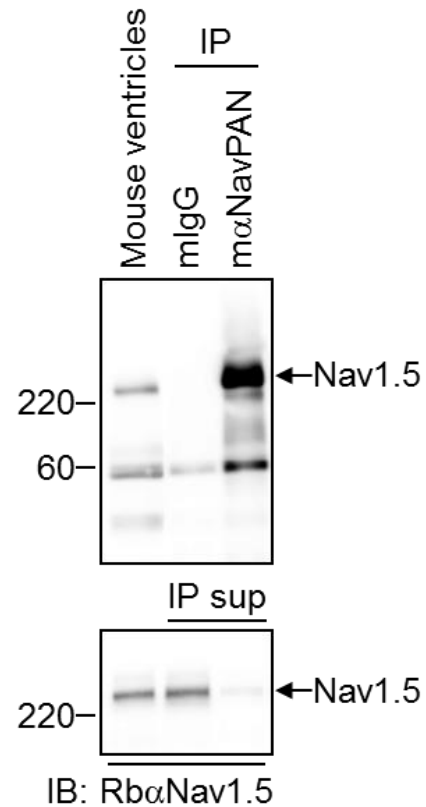
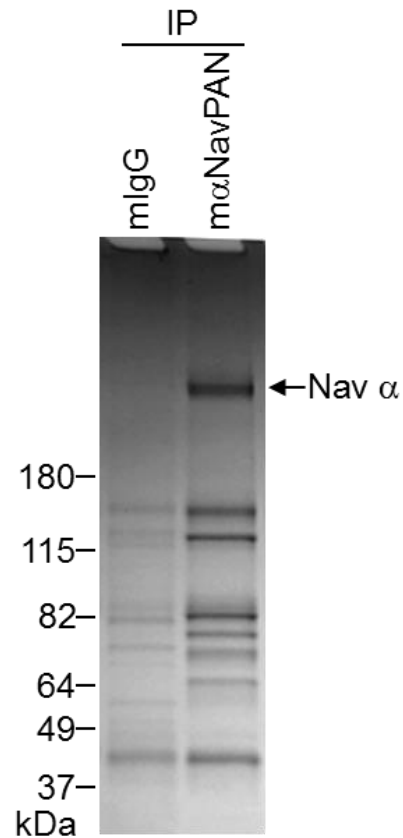
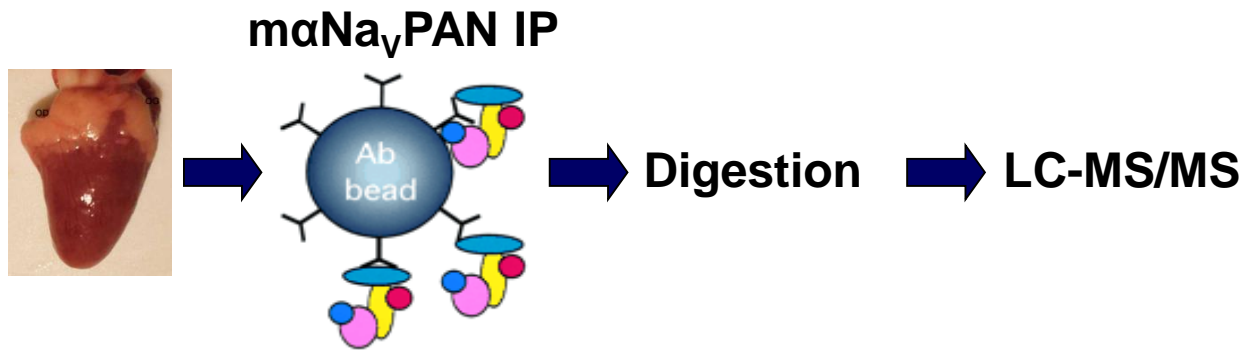
**m $\alpha$ NavPAN-IPs**



**Quantitative phosphoproteomics**

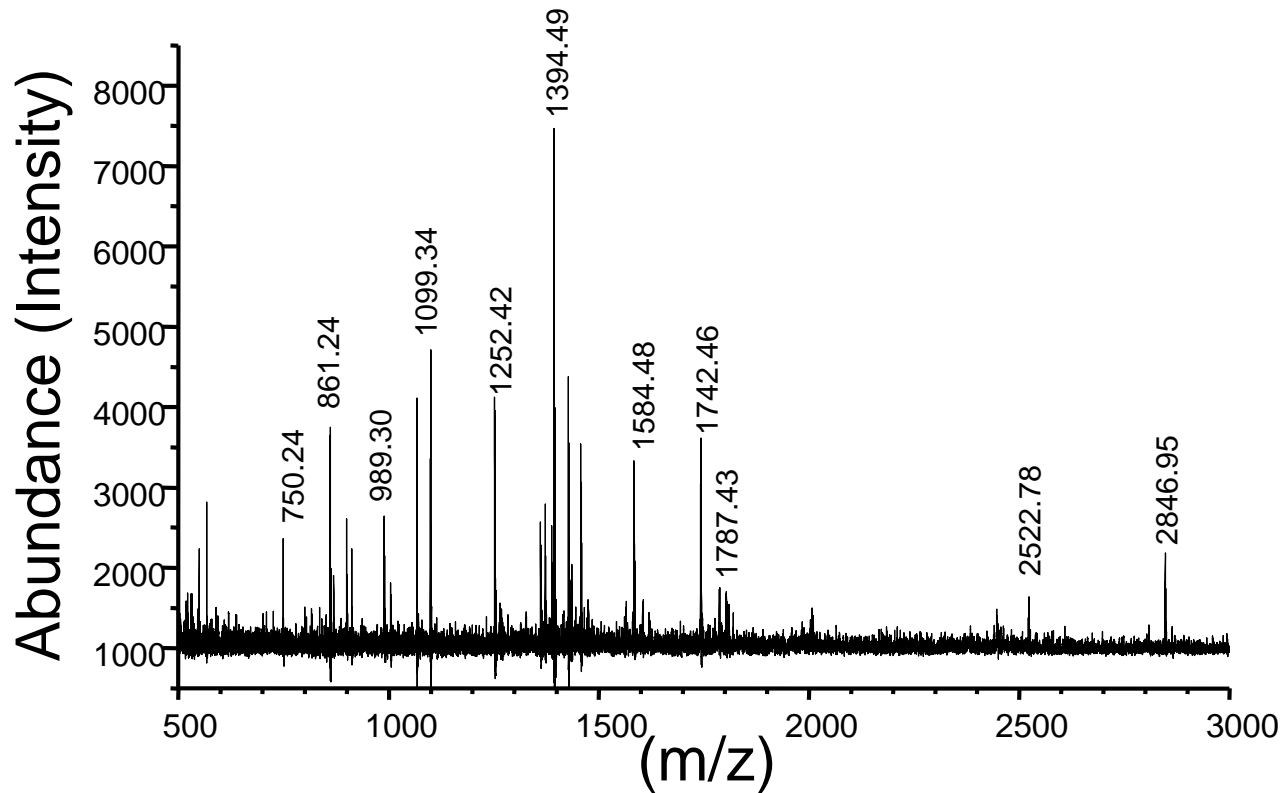


# Purification of cardiac Nav1.5 channels



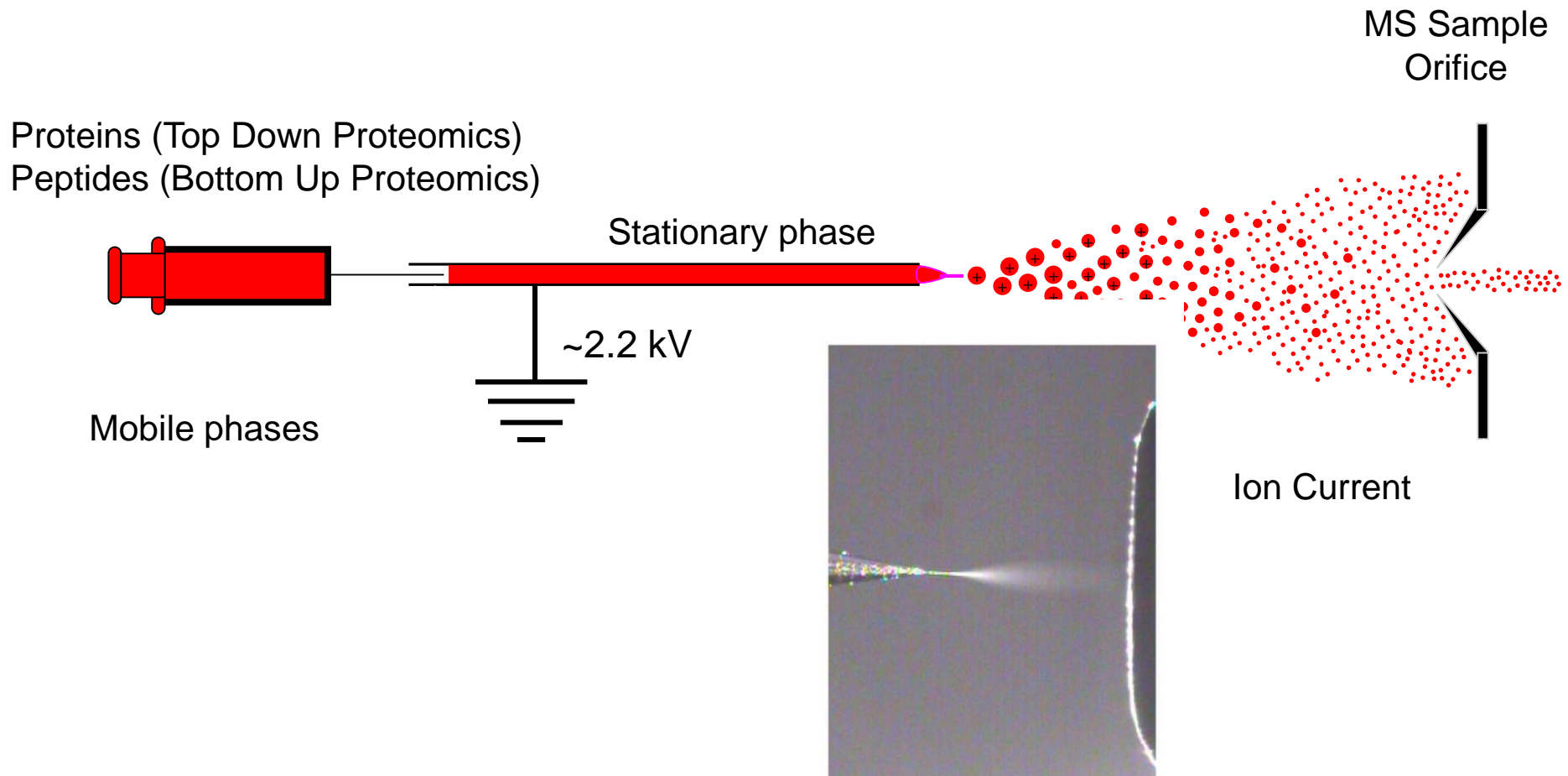
# Mass Spectrometry – A Brief Definition

Mass Spectrometry is a technique that produces charged molecules, and separates them by magnetic and/or electric fields based on their mass to charge ratio ( $m/z$ ).

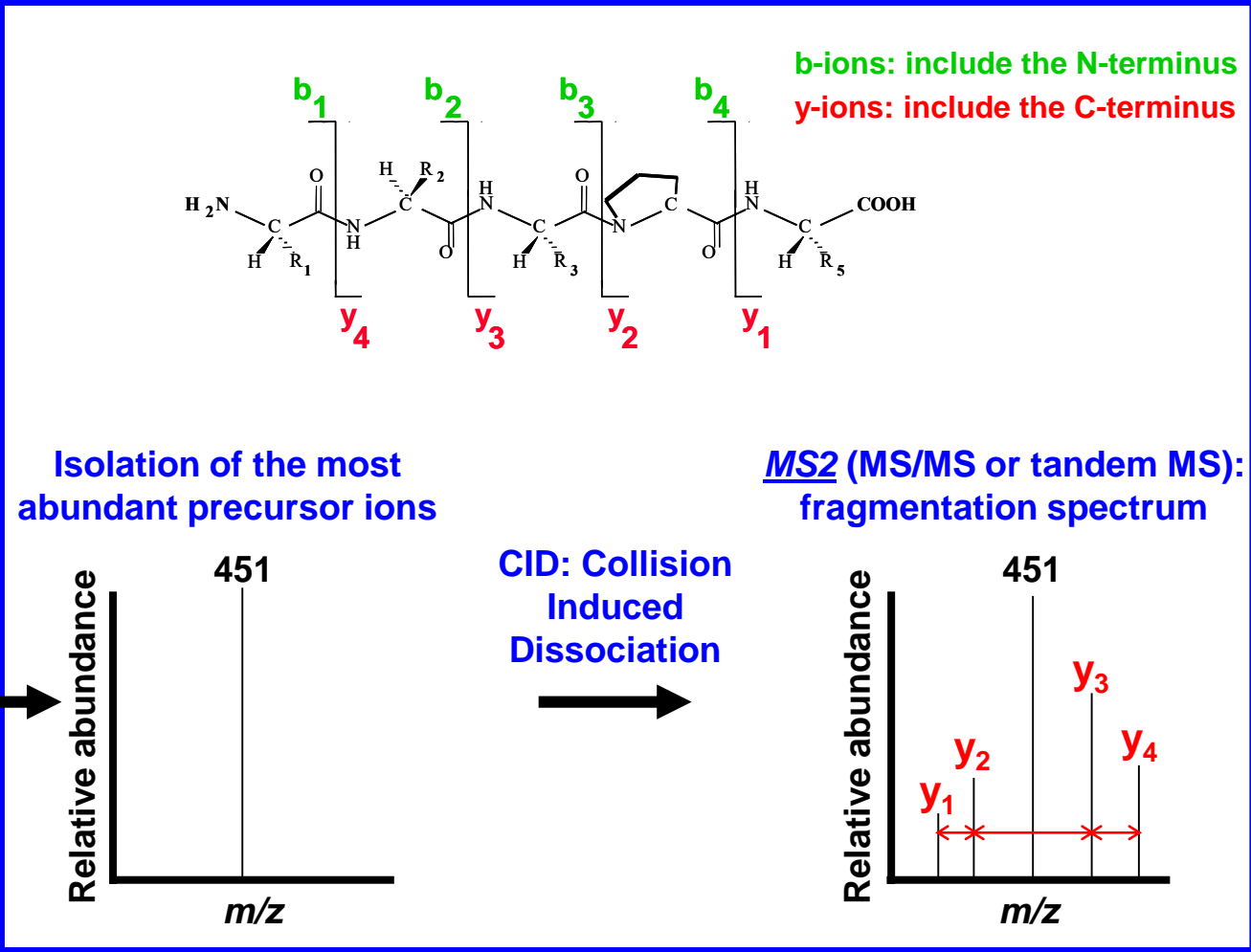
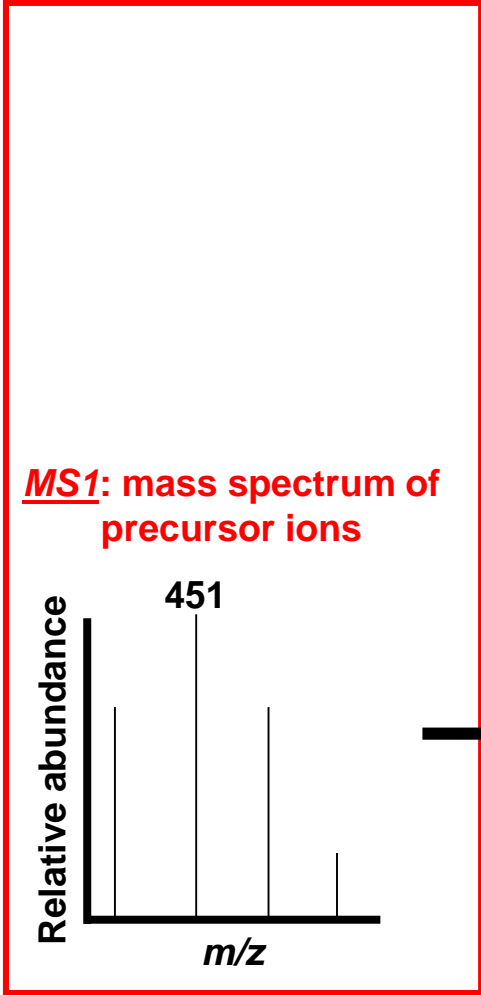


# Electrospray ionization : ESI-MS

- Couples liquid chromatography to mass spectrometry
- Reversed-phase chromatography



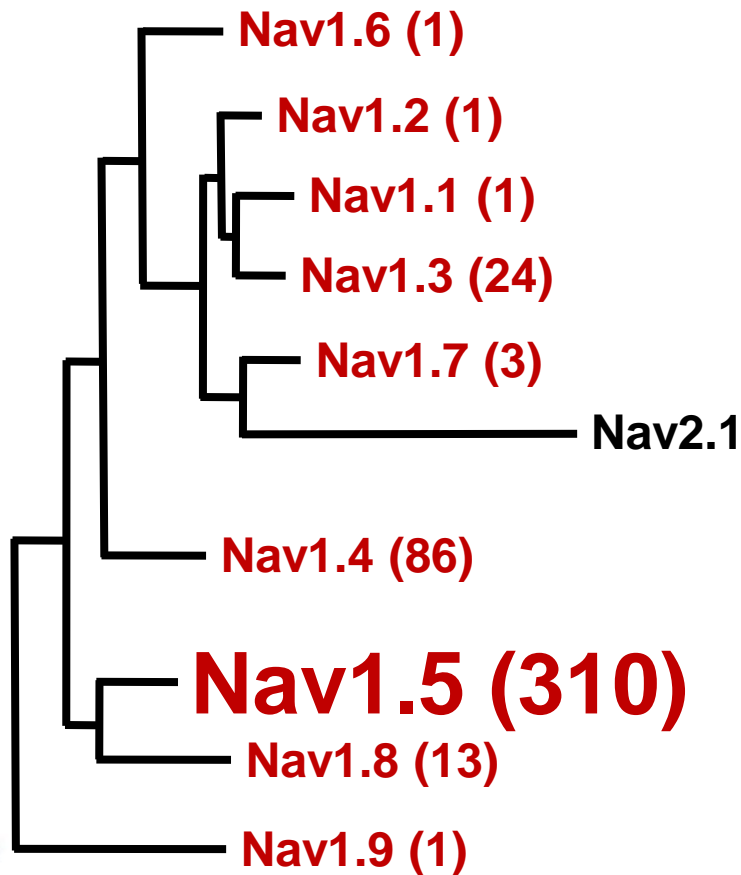
# MS/MS analyses



# Identification of Nav pore-forming subunits

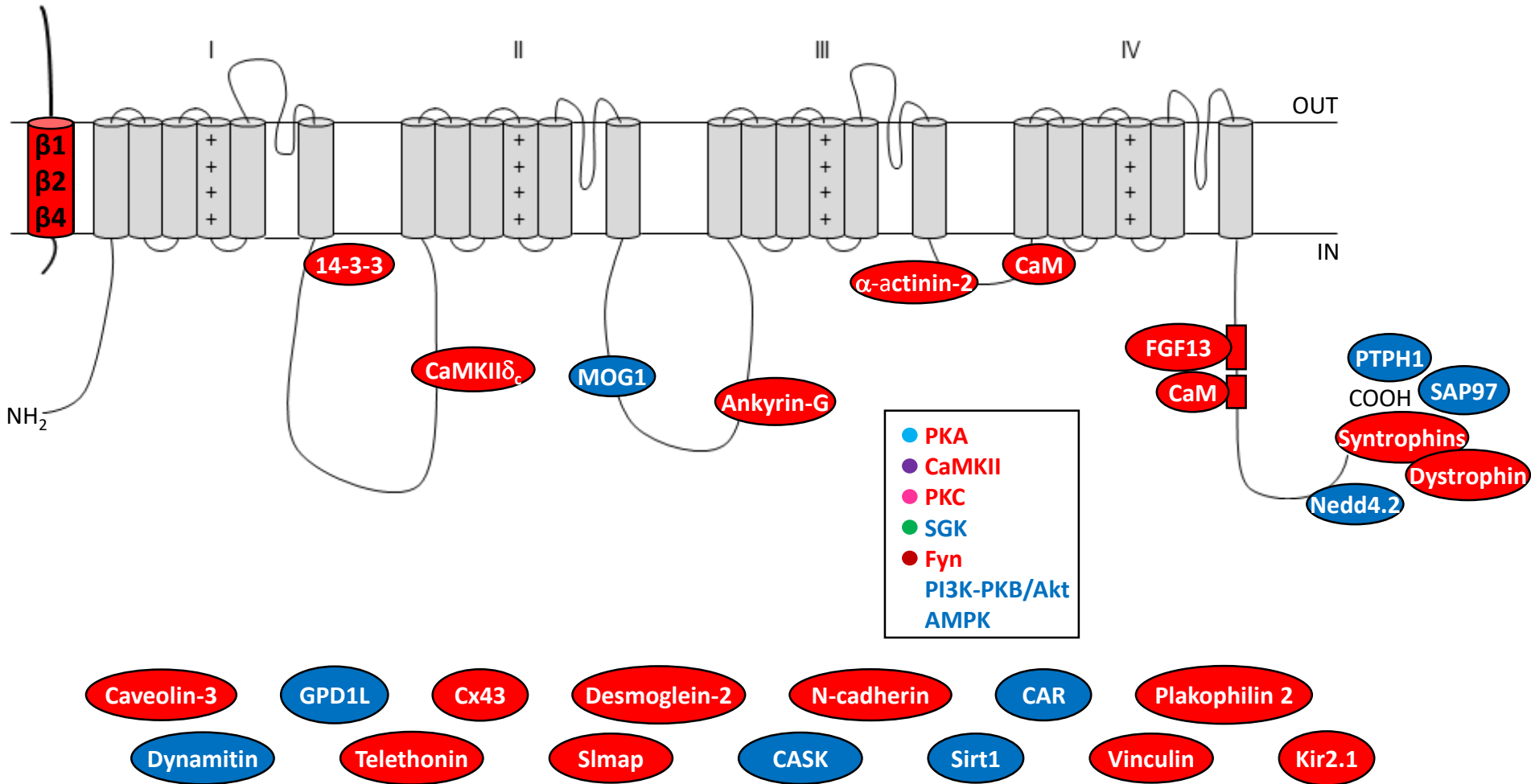
Nav<sub>v</sub>1.5 :

- Q1080 and Q1080 del isoforms
- 310 unique peptides
- 56% amino acid coverage

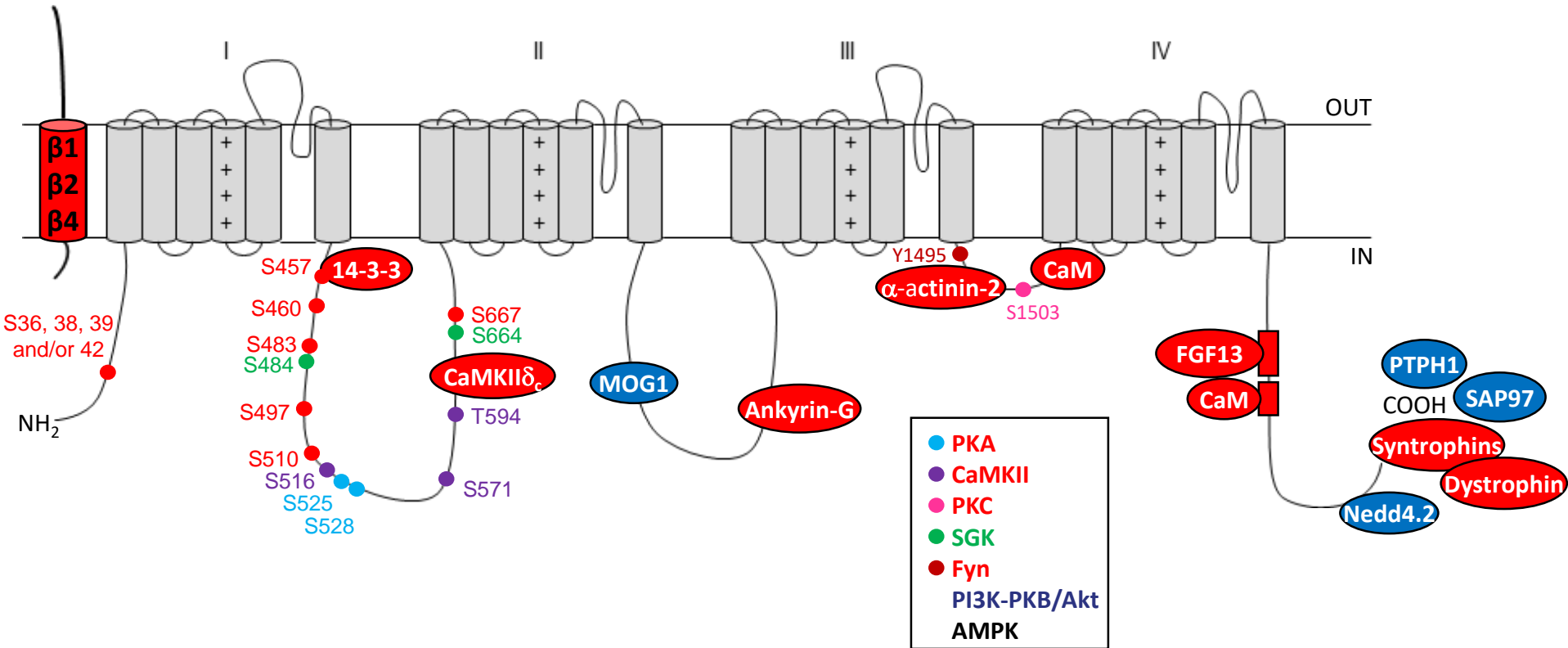


1	MANFLLPRGTS	FRR	FTRESLAAIEKR	MAEKQARCS	AT	QEE	REGLP	PEEAPRPQ	LDLQASKKLPDLYGNPPREL
	N-TERM								
76	I	G	E	P	L	E	D	L	D
	N-TERM								
151	H	D	P	P	P	T	K	Y	V
	IS2			IS3			IS4		
226	ALK	T	S	V	I	S	I	S	G
	IS5								
301	W	N	S	L	D	V	Y	L	N
	IS6								
376	R	L	Y	Q	T	L	R	S	A
	Loop I								
451	R	G	V	D	T	V	S	R	S
	Loop I								
526	R	G	S	I	F	T	F	R	R
	Loop I								
601	V	S	L	L	G	A	D	E	A
	Loop I								
676	L	E	E	S	H	R	K	C	P
	IS1								
751	V	G	N	L	V	F	T	G	I
	IS2			IS3			IS4		
826	N	T	L	I	K	I	G	N	S
	IS5								
901	W	I	E	T	M	W	D	C	M
	IS6								
976	K	R	T	T	W	D	F	C	C
	Loop II								
1051	I	A	V	A	E	S	D	D	D
	Loop II								
1124	A	E	P	R	A	P	G	C	G
	Loop II								
1199	K	T	C	Y	R	I	V	E	H
	IIIS1			IIIS2					
1274	W	C	W	L	D	F	L	I	V
	IIIS3			IIIS4					
1349	L	I	F	S	I	M	G	V	N
	IIIS5								
1424	W	M	D	I	M	A	A	V	D
	IIIS6								
1499	N	A	M	K	L	G	S	K	P
	Loop III			IVS1			IVS2		
1574	F	T	G	E	C	I	V	K	M
	IVS3			IVS4			IVS5		
1649	L	L	F	A	L	M	S	L	P
	IVS6								
1724	L	N	T	G	P	P	Y	C	D
	C-TERM								
1799	E	I	W	E	K	F	D	P	E
	C-TERM								
1874	L	K	I	Q	M	E	E	K	F
	C-TERM								
1949	E	G	L	I	A	M	N	E	N
	C-TERM								

# Characterization of cardiac Nav1.5 channel complexes



# 2012 : 9 + 7 = 16 phosphorylation sites



**Glycosylation**

**Phosphorylation**

**Méthylation**

**Acétylation**

**Oxidation**

**Nitrosylation**

**Lipoxidation**

**Ubiquitylation**

Caveolin-3

GPD1L

Cx43

Desmoglein-2

N-cadherin

CAR

Plakophilin 2

Dynamitin

Telethonin

Slmap

CASK

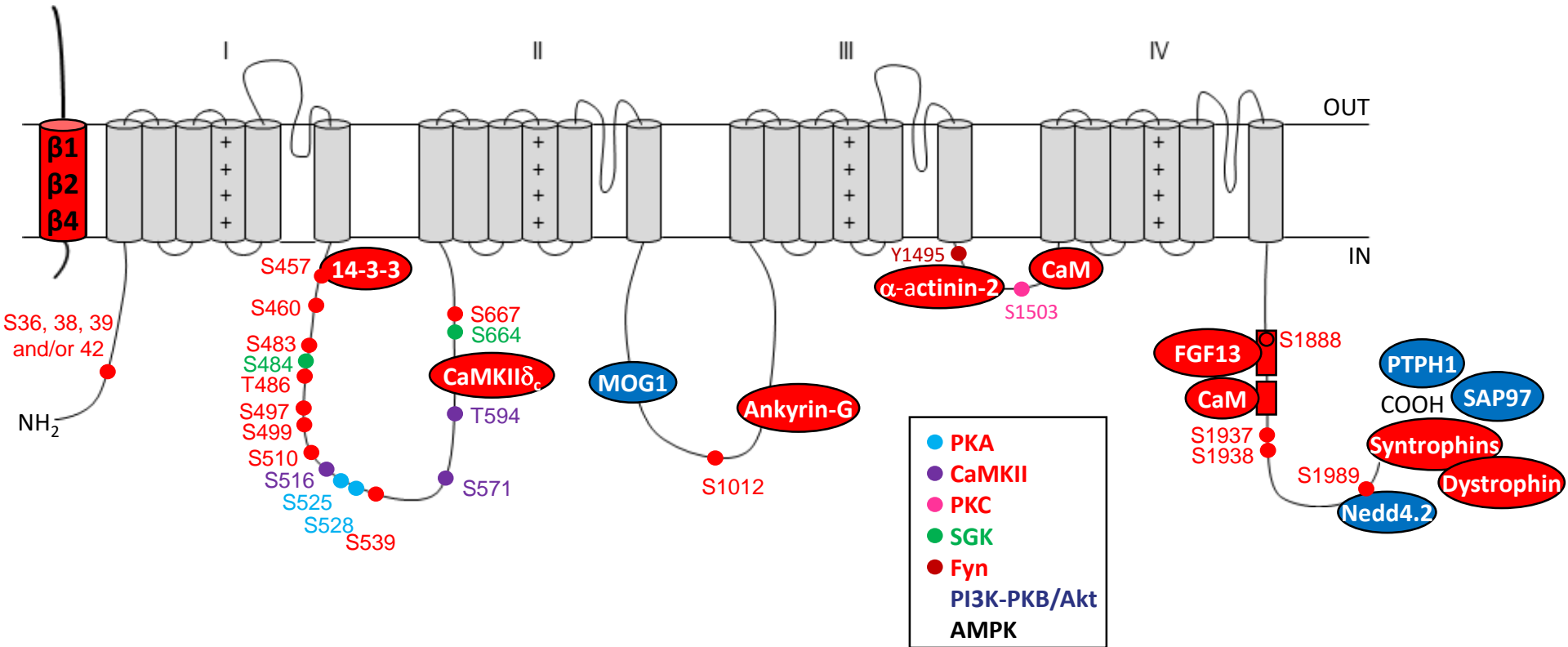
Sirt1

Vinculin

Kir2.1



# 2017 : 16 + 8 = 24 phosphorylation sites



**Glycosylation**

**Phosphorylation**

**Méthylation**

**Acétylation**

**Oxidation**

**Nitrosylation**

**Lipoxidation**

**Ubiquitylation**

**Caveolin-3**

**GPD1L**

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l'institut  
du thorax

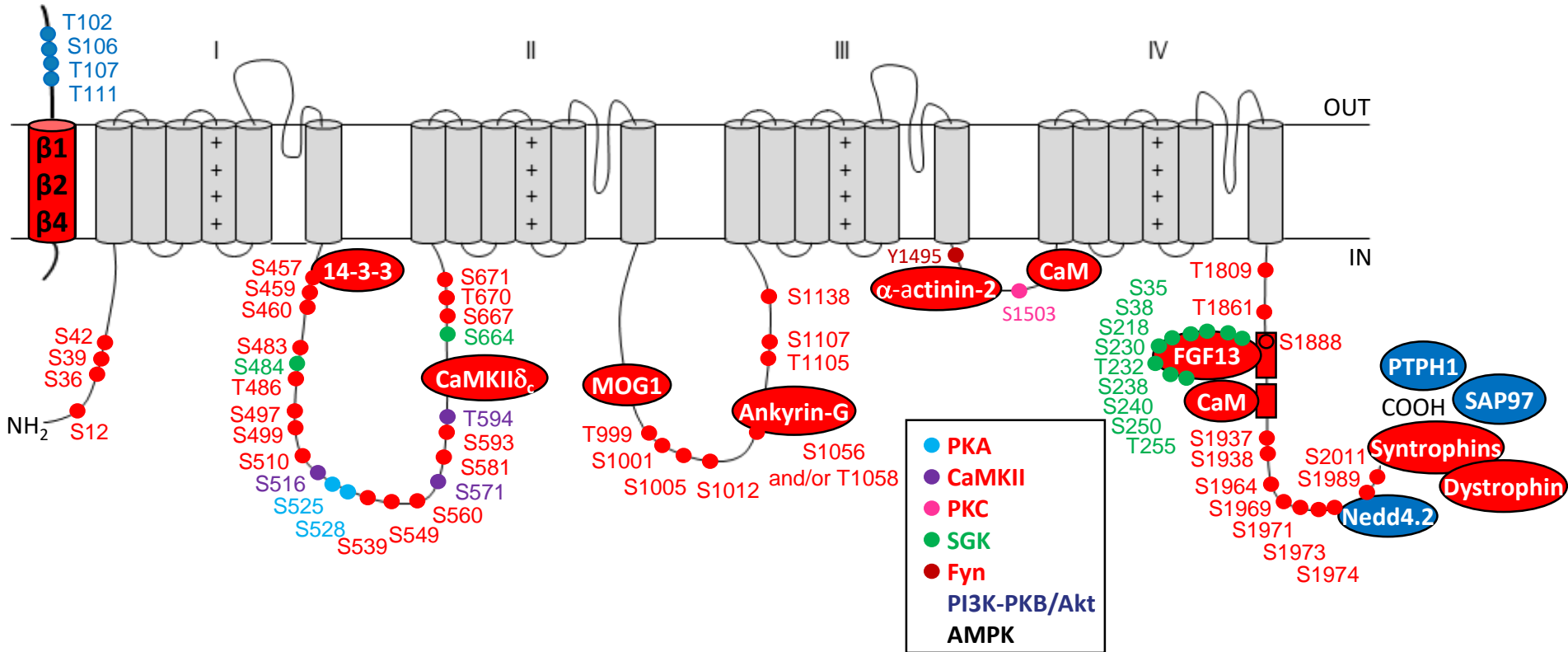
Marionneau and Abriel, *JMCC* 2015

Marionneau et al, *JPR* 2012

Burel et al, *JBC* 2017



# 2018 : 49 + 4 + 9 = 62 phosphorylation sites



**Glycosylation**

**Phosphorylation**

**Méthylation**

**Acétylation**

**Oxidation**

**Nitrosylation**

**Lipoxidation**

**Ubiquitylation**

Caveolin-3

GPD1L

Cx43

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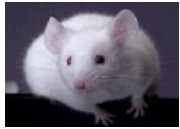
Vinculin

Kir2.1

Marionneau and Abriel, *JMCC* 2015  
 Marionneau et al, *JPR* 2012  
 Burel et al, *JBC* 2017  
 Lorenzini et al, *JGP* 2021

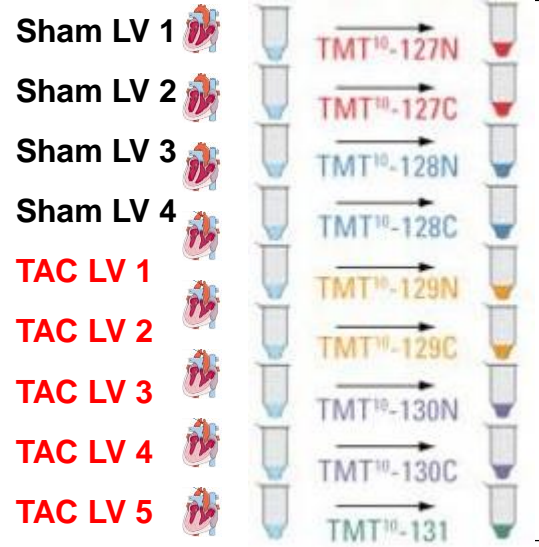
# TMT9 quantitative analysis from Sham/TAC mice

Sham / TAC mice

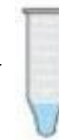


$\alpha$ Na<sub>v</sub>PAN-IPs

Labelling



Combine

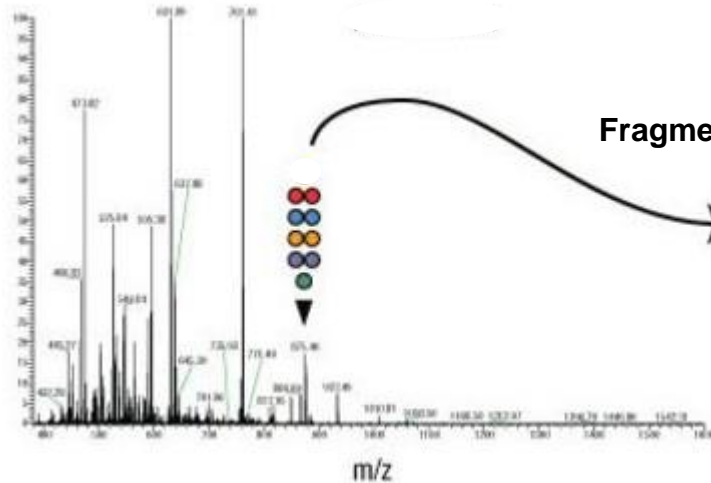


TMT methods

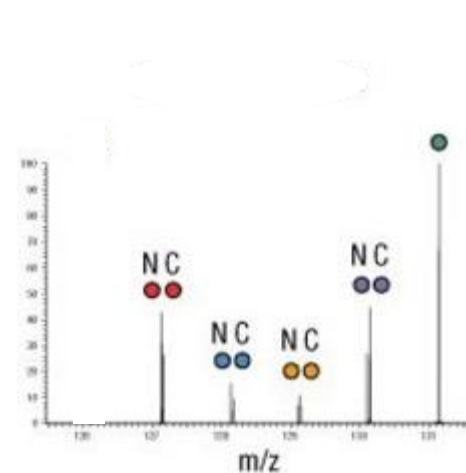
1 LC-MS/MS experiment



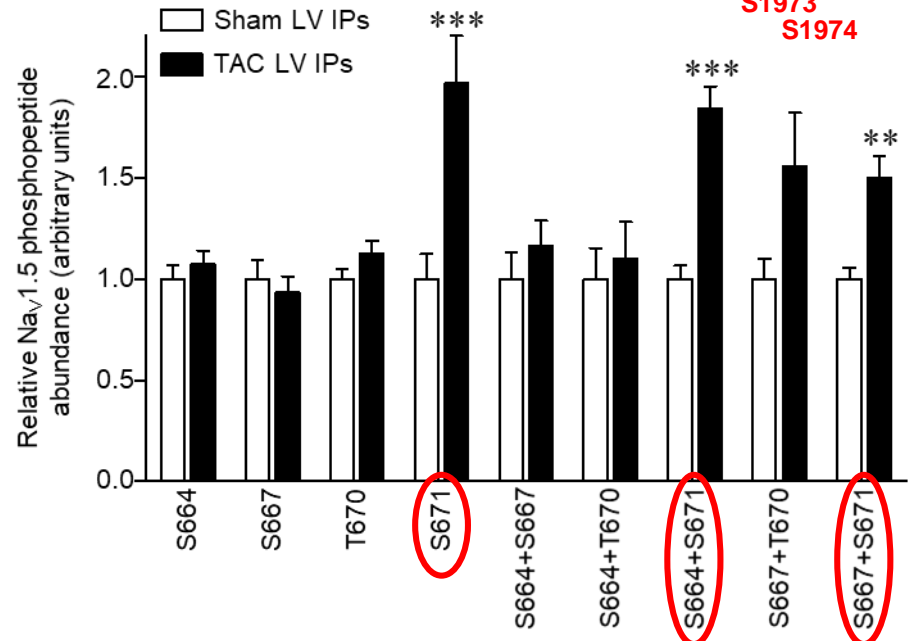
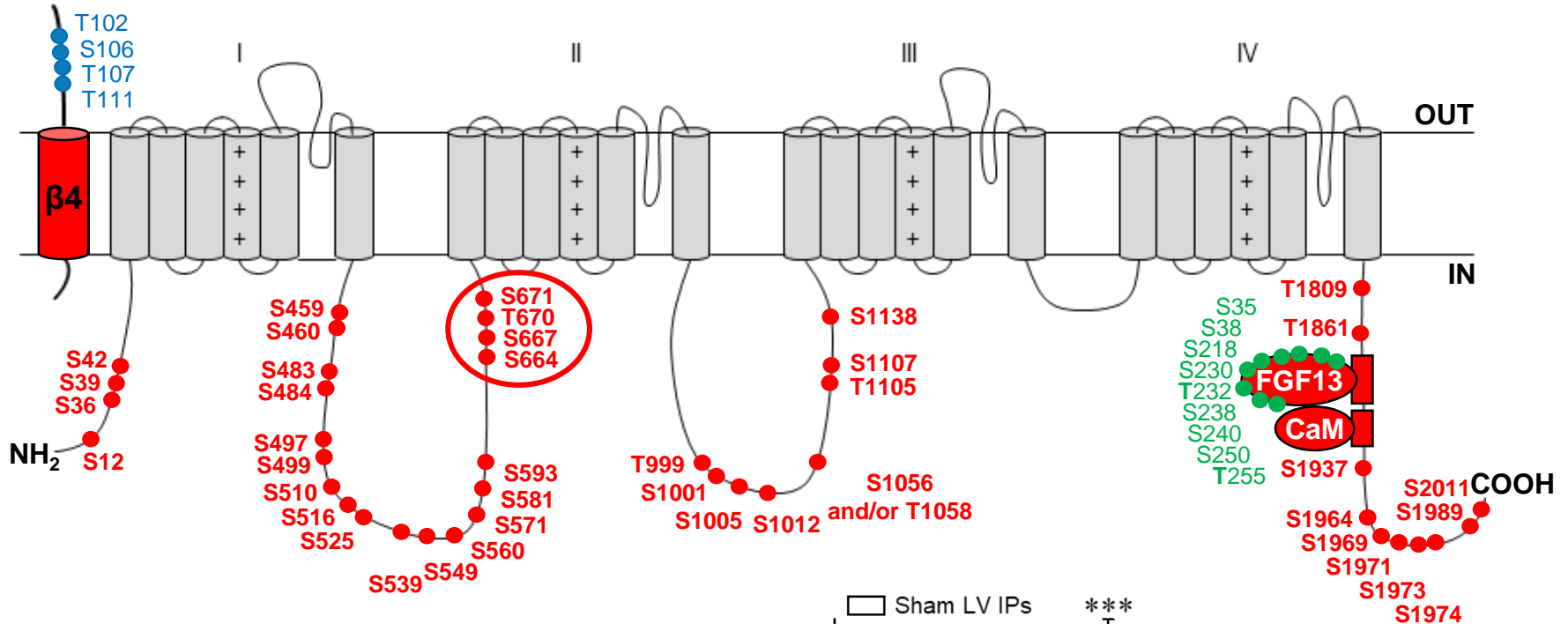
MS1 spectra: selection



MS/MS spectra: Quantification

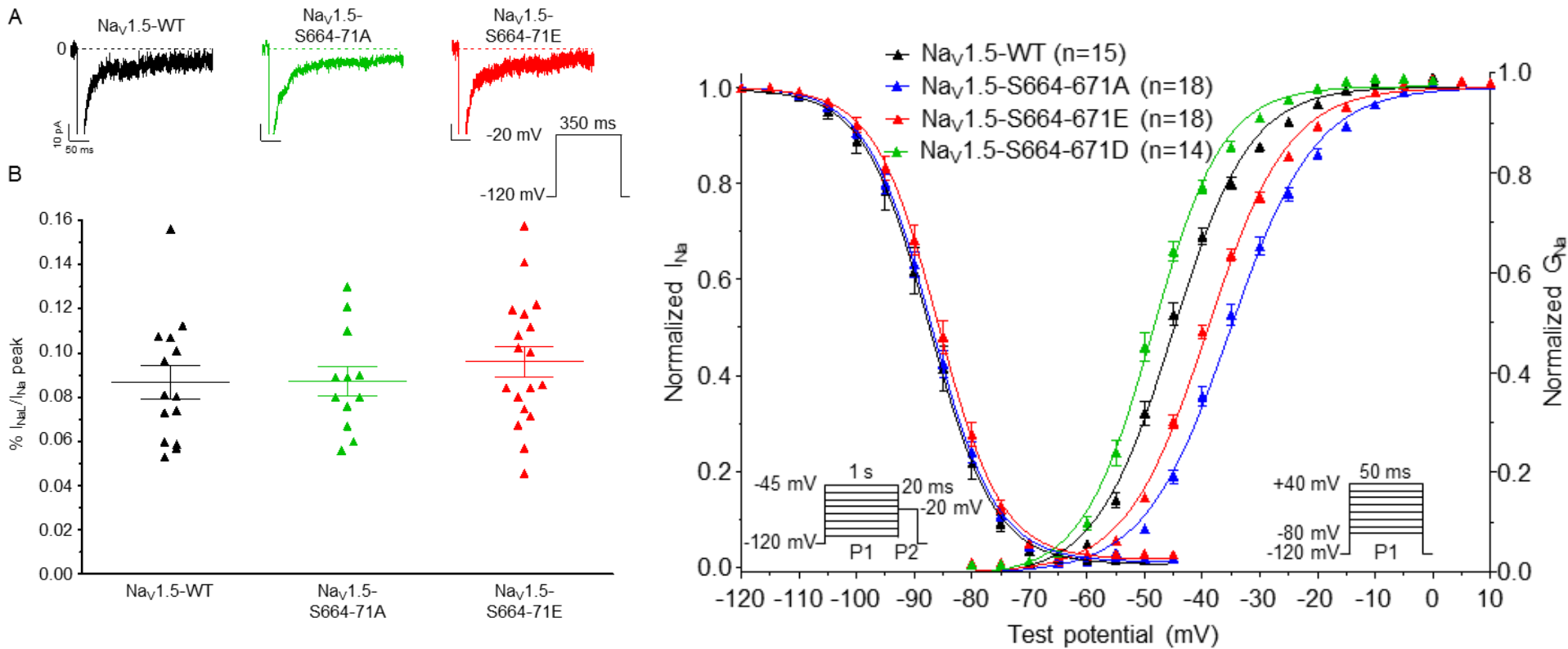


# Quantitative phosphoproteomics in Sham/TAC mice

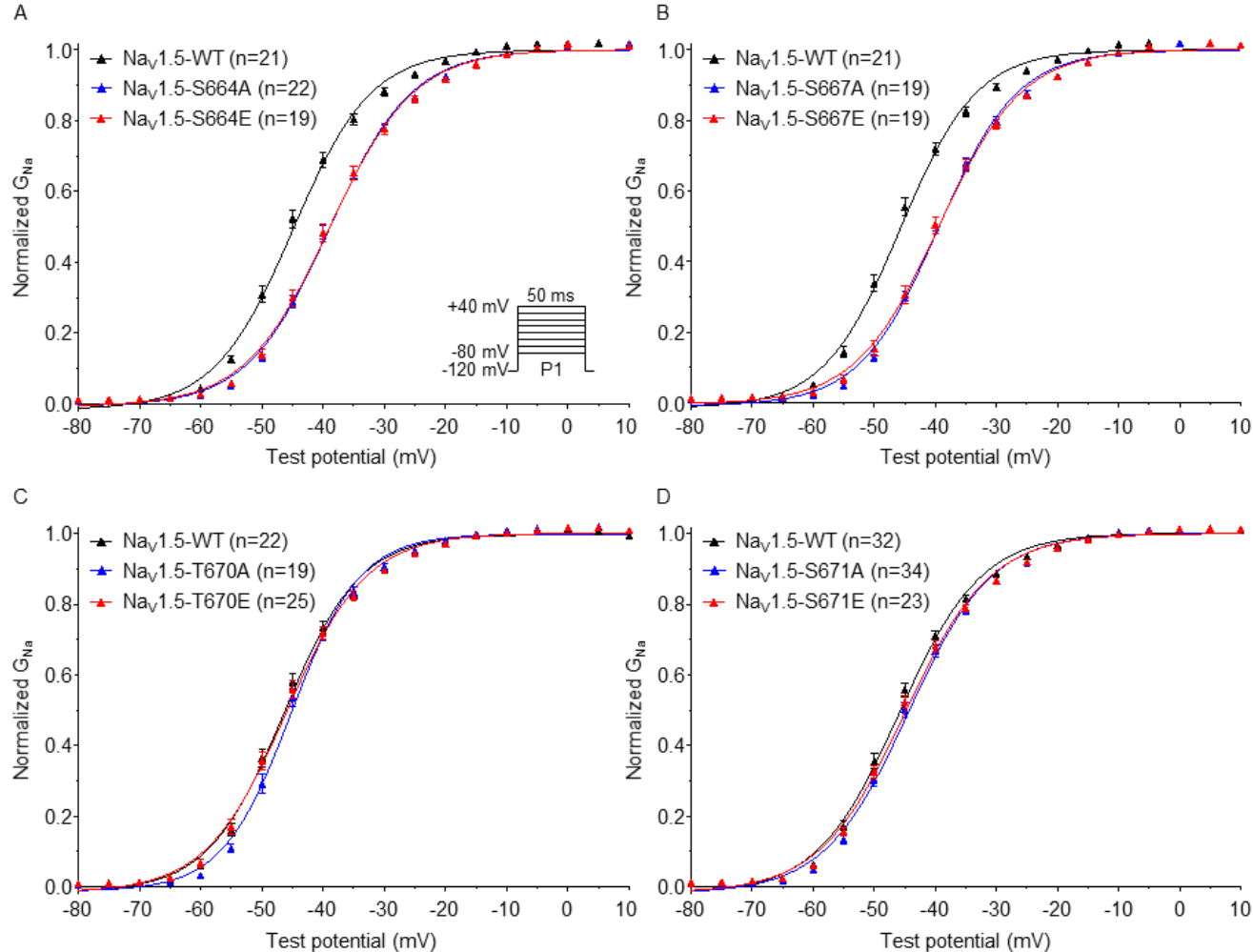


\*\*  $p < 0.01$ , \*\*\*  $p < 0.001$  in TAC vs Sham

# Electrophysiological analysis of Nav1.5-S664-671A and E mutants

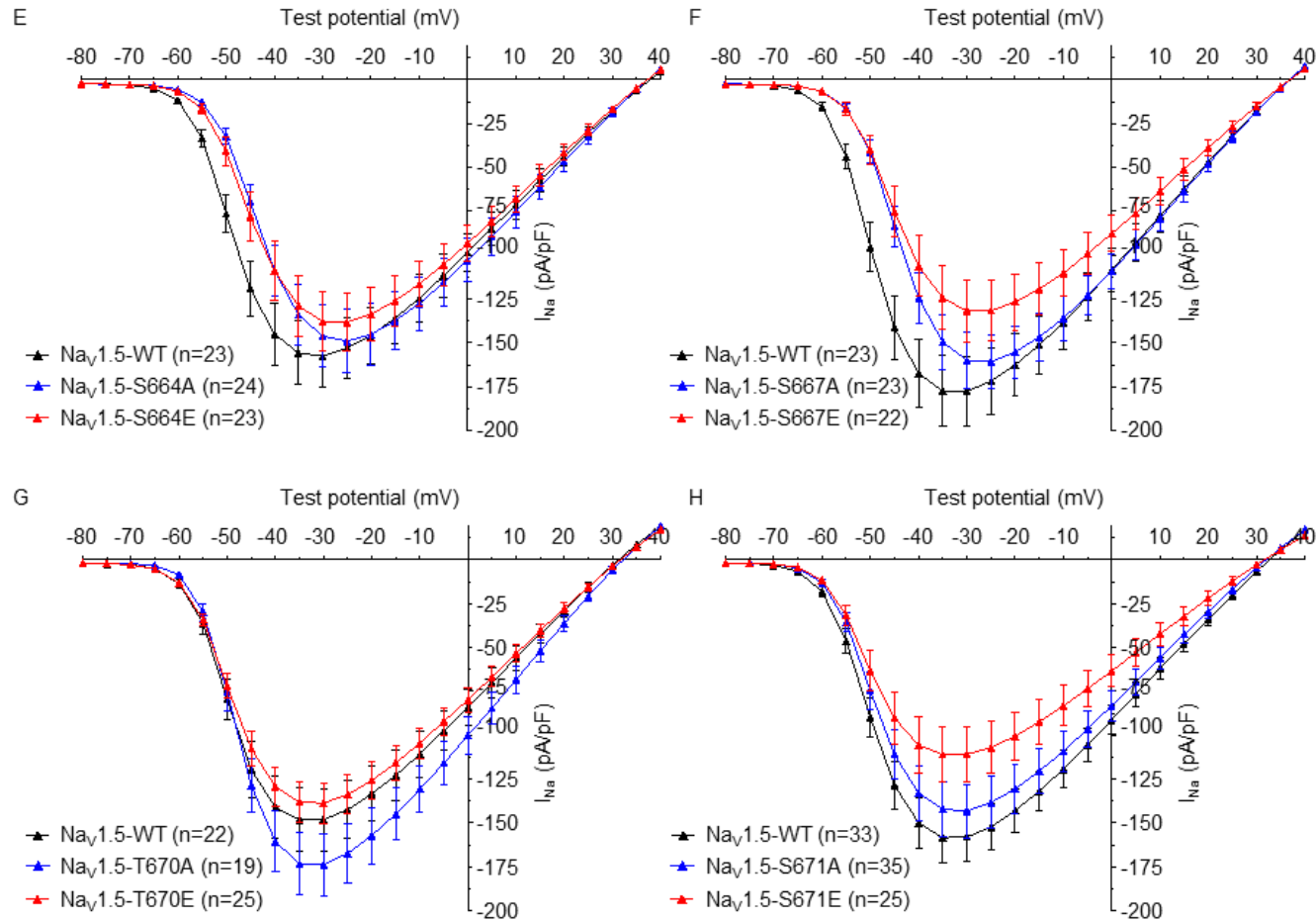


# Electrophysiological analysis of simple Nav1.5 phosphomutants



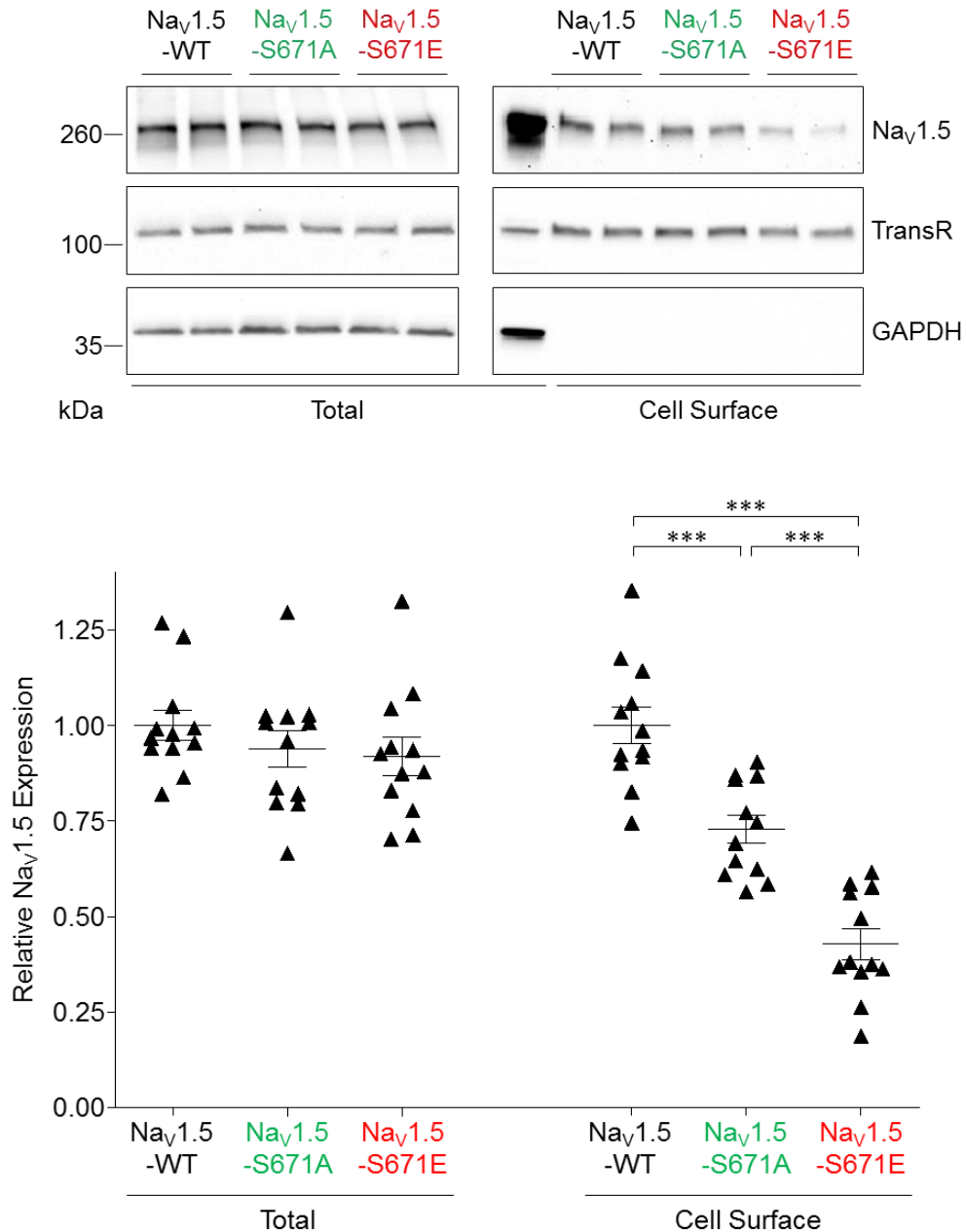
**Main finding: Phosphorylation at S664 and S667 regulates Nav1.5 channel activation.**

# Electrophysiological analysis of simple Nav1.5 phosphomutants



**Main finding: Phosphorylation at S671 decreases  $I_{Na}$ .**

# Phosphorylation at S671 decreases Nav1.5 cell surface expression



# Take-home messages

- Cardiac electrophysiology, Nav1.5 channels
- Methods to analyze macromolecular complexes and PTM
- Identification of phosphorylation sites
  - *in silico*
  - *in vitro*
  - *in situ*
- Mass spectrometry
  - How does it work?
  - Advantages *versus* other approaches



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*Flavien Charpentier*

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*Jonathan Silva*

*Reid Townsend*

**The Ohio State University, Columbus**

*Isabelle Deschènes*



**Marie Curie 7<sup>th</sup> Framework Program  
of the European Commission**

